

Ligation Method

Field of the Invention

This application relates to a method of ligating two or more molecules, for example, small organic molecules, labels, peptides etc. In particular it relates to a method of ligating a peptide, such as ligation of a synthetic peptide to a recombinant peptide.

Background to the Invention

Protein engineering methodologies have proven to be invaluable for generating protein based tools for application in basic research, diagnostics, drug discovery and as protein therapeutics. The ability to manipulate the primary structure of a protein in a controlled manner opens up many new possibilities in the biological and medical sciences. As a consequence, there is a concerted effort on developing methodologies for the site-specific modification of proteins and their subsequent application.

1

2 The two main approaches to generating proteins are
3 through recombinant methods or chemical synthesis.
4 To date, the two methods have proved to be
5 complementary; recombinant methodologies enable
6 proteins of any size to be generated but in general
7 they are restricted to the assembly of the
8 proteinogenic amino acids. Thus, in general, the
9 introduction of labels and probes into recombinant
10 proteins has to be implemented post-translationally
11 and does not allow modifications to the protein
12 backbone.

13

14 The most common methods for labelling a recombinant
15 protein use an amino or a thiol reactive version of
16 the label that will covalently react with a lysine
17 side chain / N^α amino group or a cysteine side chain
18 within the protein respectively. For such labelling
19 methods to be site-specific, an appropriate
20 derivative of the protein must be engineered to
21 contain a unique reactive functionality at the
22 position to be modified. This requires all the other
23 naturally occurring reactive functionalities within
24 the primary sequence to be removed through amino
25 acid mutagenesis. In the case of protein amino
26 functionalities, this is essentially impossible due
27 to the abundance of lysine residues within proteins
28 and the presence of the amino functionality at the
29 N-terminus of the sequence. Likewise, for cysteine
30 this process is laborious and is often detrimental
31 to the function of the protein.

32

1 The production of proteins having site-specific
2 modifications and/or labels is more readily
3 achievable using chemical synthesis methods. The
4 chemical synthesis of proteins enables multiple
5 modifications to be incorporated into both side-
6 chain and backbone moieties of the protein in a
7 site-specific manner, but, in general, the maximum
8 size of sequence that can be synthesised and
9 isolated is circa 50 - 100 amino acids.

10

11 **Protein Ligation**

12 A further approach to the generation of proteins is
13 protein / peptide ligation. In this approach
14 mutually reactive chemical functionalities
15 (orthogonal to the chemistry of the naturally
16 occurring amino acids i.e. which react by mutually
17 exclusive chemistries compared to the reactions of
18 the reactive moieties of the naturally occurring
19 amino acids) are incorporated at the N- and C-
20 termini of unprotected polypeptide fragments such
21 that when they are mixed, they react in a
22 chemoselective manner to join the two sequences
23 together (Cotton GJ and Muir TW. Chem.Biol., 1999,
24 6, R247-R254). The principle of chemical ligation is
25 shown schematically in Figure 1.

26

27 A number of chemistries have been utilised for the
28 ligation of two synthetic peptides where a diverse
29 range of different chemical functionalities can be
30 incorporated into the termini of polypeptides using
31 solid phase peptide synthesis. These include the
32 reaction between a thioacid and bromo- alkyl to

1 form a thioester (Schnolzer M and Kent SBH, Science,
2 1992, 256, 221-225), reaction of an aldehyde with an
3 N-terminal cysteine or threonine to form
4 thiazolidine or oxazolidine respectively (Liu C-F
5 and Tam J P. Proc. Natl. Acad. Sci. USA, 1994, 91,
6 6584 - 6588), reaction between a hydrazide and an
7 aldehyde to form a hydrazone (Gaertner HF et al, et
8 al Bioconj. Chem., 1992, 3, 262 - 268) reaction of
9 an aminoxy group and an aldehyde to form an oxime
10 (Rose K. J. Am. Chem. Soc., 1994, 116, 30-33),
11 reaction of azides and aryl phosphines to form an
12 amide bond (Staudinger ligation) (Nilsson BL,
13 Kiessling LL, and Raines RT. Org. Lett., 2001, 3, 9-
14 12, Kiick et al Proc. Natl. Acad. Sci. USA, 2002,
15 99, 19-24) , and the reaction of a peptide C-
16 terminal thioester and an N-terminal cysteine
17 peptide to form a native amide bond (Dawson et al.
18 Science, 1994, 266, 776) (Native chemical ligation
19 US6184344, EP 0832 096 B1). This native chemical
20 ligation method is an extension of studies by
21 Wieland and coworkers who showed that the reaction
22 of ValSPh and CysOH in aqueous buffer yielded the
23 dipeptide ValCysOH (Wieland T et al., Liebigs Ann.
24 Chem., 1953, 583, 129-149).

25

26 Although the native chemical ligation method has
27 proved popular, it requires an N-terminal cysteine
28 containing peptide for the reaction and thus, if a
29 cysteine is not present at the appropriate position
30 in the protein, a cysteine needs to be introduced at
31 the ligation site. However, the introduction of
32 extra thiol groups into a protein sequence may be

1 detrimental to its structure / function, especially
2 since cysteine has a propensity to form disulfide
3 bonds which may disrupt the folding pathway or
4 compromise the function of the folded protein.

5
6 As a consequence of the difficulties and problems
7 associated with known ligation techniques, the
8 ligation of two synthetic fragments generally only
9 enables proteins of circa 100 - 150 amino acids to
10 be chemically synthesised. Although larger proteins
11 have been synthesised by ligating together more than
12 two fragments, this has proved to be technically
13 difficult (Camarero et al. *J. Pept. Res.*, 1998, 54,
14 303-316, Canne LE et al, *J. Am. Chem. Soc.*, 1999,
15 121, 8720-8727).

16
17 **Protein semi-synthesis**

18
19 protein ligation technologies that enable both
20 synthetic and recombinantly derived protein
21 fragments to be joined together have been described.
22 This enables large proteins to be constructed from
23 combinations of synthetic and recombinant fragments,
24 allowing proteins to be site-specifically modified
25 with both natural and unnatural entities. By
26 utilising such so-called protein semi-synthesis,
27 many different synthetic moieties can be site-
28 specifically incorporated at multiple different
29 sites within a target protein.

30
31 In order to utilise recombinant proteins in ligation
32 strategies the recombinant fragments must contain

1 the appropriate reactive functionalities to
2 facilitate ligation. One approach to introduce a
3 unique reactive functionality into a recombinant
4 protein has been through the periodate oxidation of
5 N-terminal serine containing sequences. Such
6 treatment converts the N-terminal serine into a
7 glyoxyl moiety, which contains an N-terminal
8 aldehyde. Synthetic hydrazide containing peptides
9 have then been ligated to the N-terminus of these
10 proteins in a chemoselective manner through
11 hydrazone bond formation with the protein N-terminal
12 glyoxyl group (Gaertner HF et al, et al Bioconj.
13 Chem., 1992, 3, 262 - 268, Gaertner HF, et al. *J.*
14 *Biol. Chem.*, 1994, 269, 7224-7230). Another approach
15 has been to generate recombinant proteins with N-
16 terminal cysteine residues. Synthetic peptides
17 containing C-terminal thioesters have then been
18 site-specifically attached to the N-terminus of
19 these proteins via amide bond formation in a manner
20 analogous to 'native chemical ligation' (Cotton GJ
21 and Muir TW. *Chem. Biol.*, 2000, 7, 253-261). However
22 as with the ligation of synthetic peptides using
23 native chemical ligation techniques, the technology
24 requires a cysteine to be introduced at the ligation
25 site if the primary sequence does not contain one at
26 the appropriate position.

27

28 **Protein Splicing Techniques**

29

30 Recently technologies have been developed which
31 enable recombinant proteins containing C-terminal
32 thioester groups to be generated. The C-terminal

1 thioester functionality provides a unique reactive
2 chemical group within the protein that can be
3 utilised for protein ligation. Recombinant C-
4 terminal thioester proteins are produced by
5 manipulating a naturally occurring biological
6 phenomenon known as protein splicing (Paulus H. Annu
7 *Rev Biochem* 2000, 69, 447-496). Protein splicing is
8 a post-translational process in which a precursor
9 protein undergoes a series of intramolecular
10 rearrangements which result in precise removal of an
11 internal region, referred to as an intein, and
12 ligation of the two flanking sequences, termed
13 exteins (Figure 2). While there are generally no
14 sequence requirements in either of the exteins,
15 inteins are characterised by several conserved
16 sequence motifs and well over a hundred members of
17 this protein domain family have now been identified.

18

19 The first step in protein splicing involves an N→S
20 (or N→O) acyl shift in which the N-extein unit is
21 transferred to the sidechain SH or OH group of a
22 conserved Cys/Ser/Thr residue, always located at the
23 immediate N-terminus of the intein. Insights into
24 this mechanism have led to the design of a number of
25 mutant inteins which can only promote the first step
26 of protein splicing (Chong et al *Gene*. 1997, 192,
27 271-281, (Noren et al., *Angew. Chem. Int. Ed. Engl.*,
28 2000, 39, 450-466). Proteins expressed as in frame
29 N-terminal fusions to one of these engineered
30 inteins can be cleaved by thiols via an
31 intermolecular transthioesterification reaction, to
32 generate the recombinant protein C-terminal

1 thioester derivative (Figure 3) (Chong et al. *Gene*.
2 1997, 192, 271-281, (Noren et al., *Angew. Chem. Int.*
3 *Ed. Engl.*, 2000, 39, 450-466) (New England Biolabs
4 Impact System WO 00/18881, WO 0047751). Peptide
5 sequences containing an N-terminal cysteine residue
6 can then be specifically ligated to the C-termini of
7 such recombinant C-terminal thioester proteins (Muir
8 et al. *Proc. Natl. Acad. Sci. USA.*, 1998, 95, 6705-
9 6710, Evans Jr et al. *Prot. Sci.*, 1998, 7, 2256-
10 2264) , in a procedure termed expressed protein
11 ligation (EPL) or intein-mediated protein ligation
12 (IPL).

13

14 The chemoselective ligation of N-terminal cysteine
15 containing peptides to C-terminal thioester
16 containing peptides, be they synthetic or
17 recombinant, is performed typically at slightly
18 basic pH and in the presence of a thiol cofactor.
19 The strategy also requires a cysteine to be
20 introduced at the ligation site, if one is not
21 suitably positioned within the primary sequence.
22 These requirements of this ligation approach have
23 the potential to alter the structure and / or
24 function of both the protein ligation product and
25 the initial reactants.

26

27 For example, the chemokine RANTES is unstable in a
28 buffer of 100 mM NaCl, 100 mM sodium phosphate pH
29 7.4 containing 100 mM 2-mercaptoethanesulfonic acid
30 (MESNA); a buffer typically used for the ligation of
31 C-terminal thioester molecules to N-terminal
32 cysteine containing molecules (expressed protein

1 ligation and native chemical ligation). RANTES
2 contains two disulphide bonds critical for
3 maintaining the structure and function of the
4 protein. In the typical ligation buffer described
5 above, the folded protein was found to be converted
6 within 48 hours to a mixture of the reduced protein
7 and MESNA protein adducts. The majority of the
8 protein mixture subsequently formed a precipitate,
9 presumably reflecting the unfolded nature of these
10 species (Cotton, unpublished).

11

12 Accordingly, the inventors believe that ligation
13 reactions that require thiol containing buffers are,
14 in general, not suitable for maintaining the
15 integrity of disulphide bond containing proteins,
16 such as antibodies, antibody fragments and antibody
17 domains, cytokines, growth factors etc. Thus there
18 is a requirement for ligation approaches that are
19 typically performed in the absence of thiols. For
20 example, when monitored over a number of days, it
21 was found that RANTES was stable in 100 mM NaCl, 100
22 mM sodium phosphate buffer pH 7.4 and 100 mM sodium
23 acetate buffer pH 4.5 (inventor's unpublished
24 results). Ligation reactions that can be performed
25 under such conditions should therefore be applicable
26 for both disulphide and non-disulphide containing
27 proteins.

28

29 **Protein labelling**

30

31 Historically protein ligation means the joining
32 together of two peptide / protein fragments but this

1 is synonymous with protein labelling whereby the
2 label is a peptide or derivatised peptide. Equally
3 if a small non-peptidic synthetic molecule contains
4 the necessary reactive chemical functionality for
5 protein ligation, then ligation of the synthetic
6 molecule directly to either the N- or C- termini of
7 the protein affords site-specific labelling of the
8 protein. Thus technologies developed for the
9 ligation of protein fragments can also be used for
10 the direct labelling of either the N- or C- termini
11 of peptides or proteins in a site - specific manner
12 irrespective of their sequence.

13

14 Recombinant proteins containing N-terminal glyoxyl
15 functions (generated through periodate oxidation of
16 the corresponding N-terminal serine protein) have
17 been site-specific N-terminally labelled through
18 reaction with hydrazide or aminoxy derivatives of
19 the label (Geoghegan KF and Stroh JG. *Bioconj Chem.*,
20 1992, 3, 138-146, Alouni S et al. *Eur. J. Biochem.*,
21 1995, 227, 328 - 334). Also recombinant proteins
22 containing N-terminal cysteine residues have been N-
23 terminally labelled through reaction with labels
24 containing thioester functionalities, the label
25 being the acyl substituent of the thioester (Schuler
26 B and Pannell LK. *Bioconjug. Chem.*, 2002, 13, 1039-
27 43) and aldehyde functionalities (Zhao et al.
28 *Bioconj. Chem.*, 1999, 10, 424-430) to form amides
29 and thiazolidines respectively.

30

31 Though a number of methods for ligation of proteins
32 exist each one has its potential drawbacks. There

1 is thus a need for novel ligation methodologies,
2 especially those that are compatible with both
3 synthetic and recombinant fragments, and which may
4 be used in the ligation of disulphide bond
5 containing proteins as well as non disulphide bond
6 containing proteins, which will complement the
7 existing technologies and add another string to the
8 protein engineer's bow.

9

10 **Summary of the Invention**

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12 The present inventors have overcome a number of
13 problems associated with the prior art and have
14 developed a new method for ligating peptide
15 molecules which overcomes a number of the problems
16 of the prior art.

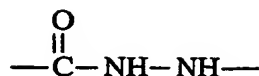
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18 Accordingly, in a first aspect of the present
19 invention, there is provided a method of producing
20 an oligopeptide product, the method comprising the
21 steps:

- 22 a) providing a first oligopeptide, the first
- 23 oligopeptide having a reactive moiety,
- 24 b) providing a second oligopeptide, the second
- 25 oligopeptide having an activated ester moiety
- 26 c) allowing the reactive moiety of the first
- 27 oligopeptide to react with the activated ester
- 28 moiety of the second oligopeptide to form an
- 29 oligopeptide product, in which the first and second
- 30 oligopeptides are linked via a linking moiety having
- 31 Formula I, Formula II or Formula III.

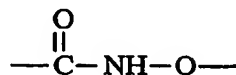
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1 Formula I



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3 Formula II



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5 Formula III



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9 In preferred embodiments, in step (c), where said
10 oligopeptides are linked via a linking moiety having
11 Formula II and where said activated ester moiety of
12 step (b) is not a thioester, said activated ester is
13 a terminal activated ester moiety.

14

15 In further preferred embodiments of the invention,
16 said linking moieties are linked via a linking
17 moiety having Formula I or Formula III.

18

19 Unless the context demands otherwise, the terms
20 peptide, oligopeptide, polypeptide and protein are
21 used interchangeably.

22

23 The activated ester moiety of the first oligopeptide
24 may be any suitable activated ester moiety, such as
25 a thioester moiety, a phenolic ester moiety, an

1 hydroxysuccinimide moiety, or an O-acylisourea
2 moiety.

3

4 In preferred embodiments of the invention, the
5 activated ester moiety is a thioester moiety. Any
6 suitable thioester peptides wherein the peptide is
7 the acyl substituent of the thioester may be used in
8 the present invention (Figure 4).

9

10 Such thioester peptides may be synthetically or
11 recombinantly produced. The skilled person is well
12 aware of methods known in the art for generating
13 synthetic peptide thioesters. For example, synthetic
14 peptide thioesters may be produced via synthesis on
15 a resin that generates a C-terminal thioester upon
16 HF cleavage (Hojo et al, Bull. Chem. Soc. Jpn.,
17 1993, 66, 2700-2706). Further, the use of 'safety
18 catch' linkers has proved to be popular for
19 generating C-terminal thioesters through thiol
20 induced resin cleavage of the assembled peptide
21 (Shin Y et al, J. Am. Chem. Soc., 1999, 121, 11684-
22 11689).

23

24 Moreover, recently technologies have been developed
25 which enable recombinant C-terminal thioester
26 proteins to be generated. Recombinant C-terminal
27 thioester proteins may be produced by manipulating a
28 naturally occurring biological phenomenon known as
29 protein splicing. As described above, protein
30 splicing is a post-translational process in which a
31 precursor protein undergoes a series of
32 intramolecular rearrangements which result in

1 precise removal of an internal region, referred to
2 as an intein, and ligation of the two flanking
3 sequences, termed exteins.

4
5 As described above, a number of mutant inteins which
6 can only promote the first step of protein splicing
7 have been designed (Chong et al *Gene*. 1997, 192,
8 271-281, Noren et al., *Angew. Chem. Int. Ed. Engl.*,
9 2000, 39, 450-466). Proteins expressed as in frame
10 N-terminal fusions to one of these engineered
11 inteins can be cleaved by thiols via an
12 intermolecular transthioesterification reaction, to
13 generate the recombinant protein C-terminal
14 thioester derivative (Chong et al *Gene*. 1997, 192,
15 271-281, Noren et al., *Angew. Chem. Int. Ed. Engl.*,
16 2000, 39, 450-466) (New England Biolabs Impact
17 System WO 00/18881, WO 0047751). Such protein
18 thioesters may be used in the methods of the
19 invention (See Figure 3).

20
21 Accordingly, in a preferred aspect of the present
22 invention, in step (b), the second oligopeptide is
23 generated by thiol reagent induced cleavage of an
24 intein fusion protein.

25
26 Accordingly, in a second aspect of the present
27 invention, there is provided a method of producing
28 an oligopeptide product, the method comprising the
29 steps:

30 a) providing a first oligopeptide, the first
31 oligopeptide having a reactive moiety,

1 b) (i) providing a precursor oligopeptide
2 molecule, the precursor oligopeptide molecule
3 comprising a precursor second oligopeptide fused N-
4 terminally to an intein domain
5 (ii) allowing thiol reagent dependent cleavage of
6 the precursor molecule to generate a second
7 oligopeptide molecule, said second oligopeptide
8 molecule having a thioester moiety at its C-terminus
9 c) allowing the reactive moiety of the first
10 oligopeptide to react with the second oligopeptide
11 molecule to form an oligopeptide product, in which
12 the first and second oligopeptides are linked via a
13 linking moiety having Formula I, II or III.

14
15 The reactive moiety of the first oligopeptide may be
16 any suitable reactive moiety. In preferred
17 embodiments of the invention, the reactive moiety is
18 a hydrazine moiety, an amino-oxy moiety or a
19 hydrazide moiety having general formula IV, V or VI
20 respectively.

21
22 Formula IV

23 —NH—NH₂

24

25

26 Formula V

—O—NH₂

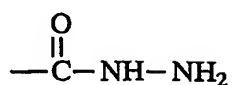
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30 Formula VI

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For example, in a particular preferred embodiment, the reactive moiety has Formula IV and, in the oligopeptide product produced by the method of the invention, the first and second oligopeptides are linked via a linking moiety having Formula I.

In a further preferred embodiment, the reactive moiety has Formula V and, in the oligopeptide product produced by the method of the invention, the first and second oligopeptides are linked via a linking moiety having Formula II.

In another preferred embodiment, the reactive moiety has Formula VI and, in the oligopeptide product produced by the method of the invention, the first and second oligopeptides are linked via a linking moiety having Formula III.

As described above, the first oligopeptide comprises a reactive moiety, which, in preferred embodiments, may be a hydrazine moiety (e.g. Formula IV), an amino-oxy moiety (e.g. Formula V) or an hydrazide moiety (e.g. Formula VI).

A particular advantage of the ligation method of the invention is that it may be performed in the absence of thiols. This enables efficient ligation of proteins/peptides comprising disulphide bonds as well as of proteins without such bonds.

1 Accordingly, in an embodiment of the first and
2 second aspects of the invention, at least one of the
3 first and second oligopeptides comprises one or more
4 disulphide bonds.

5

6 Hydrazine, hydrazide or aminooxy containing
7 derivatives of synthetic oligopeptides may be
8 readily produced using known methods, for example,
9 solid phase synthesis techniques.

10

11 Further, the present inventors have also found that
12 proteins fused N-terminal to an intein domain can be
13 cleaved from the intein by hydrazine treatment in a
14 selective manner to liberate the desired protein as
15 its corresponding hydrazide derivative (for example,
16 see Figure 5).

17

18 Accordingly, in further preferred embodiments of the
19 invention, the first oligopeptide is generated by
20 reaction of hydrazine with an oligopeptide molecule
21 comprising the first oligopeptide fused N-terminal
22 to an intein domain.

23

24 Indeed the discovery that such protein hydrazides
25 may be produced by means of such a reaction forms an
26 independent aspect of the present invention.

27

28 Accordingly, a third aspect of the invention
29 provides a method of generating a protein hydrazide,
30 said method comprising the steps:

31 (a) providing an protein molecule comprising an
32 oligopeptide fused N-terminal to an intein domain,

1 (b) reacting said protein molecule with hydrazine,
2 such that the intein domain is cleaved from the
3 oligopeptide to generate a protein hydrazide.

4
5 Moreover, as well as using such a reaction to
6 generate a first oligopeptide having a hydrazide
7 moiety at its C-terminal, the first oligopeptide
8 thus being available for reaction with the second
9 oligopeptide having the activated ester moiety, the
10 present invention further extends to a "one-step"
11 process for ligating two peptides to generate an
12 oligopeptide product.

13
14 This may be achieved by reacting a suitable protein
15 linked N-terminal to an intein directly with a
16 polypeptide having a hydrazine, hydrazide or amino-
17 oxy moiety.

18
19 Accordingly, in a fourth aspect, the invention
20 provides a method of producing an oligopeptide
21 product, the method comprising the steps:

22 a) providing a first oligopeptide, the first
23 oligopeptide having a reactive moiety, wherein the
24 reactive moiety is a hydrazine moiety, a hydrazide
25 moiety or an amino-oxy moiety;
26 (i) providing a precursor oligopeptide molecule, the
27 precursor oligopeptide molecule comprising a second
28 oligopeptide fused N-terminally to an intein domain;
29 (c) allowing the reactive moiety of the first
30 oligopeptide to react with the precursor
31 oligopeptide molecule to form an oligopeptide
32 product, in which the first and second oligopeptides

1 are linked via a linking moiety having Formula I,
2 Formula II or Formula III.

3
4 The ligation technology of the present invention can
5 thus utilise both synthetic and recombinant proteins
6 and peptides. It thus enables the ligation of two or
7 more synthetic peptides, the ligation of two or more
8 recombinant peptides or the ligation of at least one
9 synthetic peptide with at least one recombinant
10 peptide.

11
12 Moreover, as well as providing a novel method of
13 ligating peptides, the present invention may be used
14 for the labelling of synthetic or recombinant
15 peptides.

16
17 Accordingly, in a fifth aspect of the present
18 invention, there is provided a method of labelling
19 an oligopeptide, the method comprising the steps:
20 a) providing a label molecule, the label molecule
21 having a reactive moiety,
22 b) providing the oligopeptide, the oligopeptide
23 having an activated ester moiety
24 c) allowing the reactive moiety of the label
25 molecule to react with the activated ester moiety of
26 the oligopeptide to form the labelled oligopeptide,
27 in which the label molecule and the oligopeptide are
28 linked via a linking moiety having Formula I,
29 Formula II or Formula III as defined above,

30
31 In preferred embodiments, in step (c), where said
32 label molecule and the oligopeptide are linked via a

1 linking moiety having Formula II and where said
2 activated ester moiety of step (b) is not a
3 thioester, said activated ester is a terminal
4 activated ester moiety.
5

6 In a preferred aspect of the present invention, in
7 step (b) the oligopeptide is generated by thiol
8 induced cleavage of an intein fusion protein.
9

10 Accordingly, in a sixth aspect of the present
11 invention, there is provided a method of labelling
12 an oligopeptide, the method comprising the steps:
13 a) providing a label molecule, the label molecule
14 having a reactive moiety,
15 c) (i) providing a precursor oligopeptide
16 molecule, the precursor oligopeptide molecule
17 comprising a precursor oligopeptide fused N-
18 terminally to an intein domain
19 (ii) allowing thiol reagent dependent cleavage of
20 the precursor molecule to generate an oligopeptide
21 molecule, said oligopeptide molecule having a
22 thioester moiety at its C-terminus
23 c) allowing the reactive moiety of the label
24 molecule to react with the oligopeptide to form the
25 labelled oligopeptide, in which the label molecule
26 and the oligopeptide are linked via a linking moiety
27 having Formula I, II or III.
28

29 Alternatively, a label molecule having a terminal
30 activated ester moiety may be used to label an
31 oligopeptide having a reactive moiety. Thus, in a
32 seventh aspect of the invention, there is provided a

1 method of labelling an oligopeptide, the method
2 comprising the steps:
3 a) providing a label molecule, the label molecule
4 having an activated ester moiety of which the label
5 is the acyl substituent,
6 b) providing the oligopeptide, the oligopeptide
7 having a reactive moiety
8 c) allowing the activated ester moiety of the label
9 molecule to react with the reactive moiety of the
10 oligopeptide to form the labelled oligopeptide, in
11 which the label molecule and the oligopeptide are
12 linked via a linking moiety having Formula I,
13 Formula II or Formula III

14 wherein, in step (c), where said label molecule
15 and the oligopeptide are linked via a linking moiety
16 having Formula II and where said activated ester
17 moiety of step (b) is not a thioester, said
18 activated ester is a terminal activated ester
19 moiety.

20

21 As with the ligation technology, an oligopeptide
22 present as a precursor molecule linked to an intein
23 molecule may be labelled directly. Thus, an eighth
24 aspect of the present invention provides a method of
25 labelling an oligopeptide, the method comprising the
26 steps:

27 a) providing a label molecule, the label molecule
28 having a reactive moiety,
29 b) providing a precursor oligopeptide molecule,
30 the precursor oligopeptide molecule comprising an
31 oligopeptide fused N-terminally to an intein domain,

1 c) allowing the reactive moiety of the label
2 molecule to react with the precursor oligopeptide
3 molecule to form a labelled oligopeptide product, in
4 which the label molecule and the oligopeptide are
5 linked via a linking moiety having Formula I,
6 Formula II or Formula III as defined above.

7
8 Any suitable label molecule known to the skilled
9 person may be used in methods of the invention. The
10 choice of label will depend on the use to which the
11 labelled peptide is to be put. For example labels
12 which may be used in the methods of the invention
13 may include fluorophores, crosslinking reagents,
14 spin labels, affinity probes, imaging reagents, for
15 example radioisotopes, chelating agents such as
16 DOTA, polymers such as PEG, lipids, sugars, cytotoxic
17 agents, and solid surfaces and beads.

18
19 In particular embodiments of the fifth, sixth, and
20 seventh aspects of the invention, at least one of
21 the label and oligopeptides comprises one or more
22 disulphide bonds.

23
24 The methods of the invention are particularly useful
25 in the ligation of peptides, in particular the
26 ligation of peptides, which, using conventional
27 ligation techniques, would require various
28 protecting groups. The inventors have shown that
29 the methods of the invention may be performed under
30 pH conditions in which only the reactive moieties
31 will react.

32

1 In preferred embodiments of the first and second and
2 in preferred embodiments of the fourth to eighth
3 aspects of the invention, step (c) of the method is
4 performed at a pH in the range pH 4.0 to pH 8.5,
5 preferably pH 4.0 to 8.0, for example, pH 4.0 to
6 7.5, more preferably in the range pH 5.0 to pH 8.0,
7 more preferably in the range pH 6.0 to pH 7.5, most
8 preferably in the range pH 6.5 to pH 7.5.

9
10 For example, the inventors have demonstrated that
11 synthetic peptide C-terminal thioesters specifically
12 react with hydrazine under aqueous conditions at pH
13 6.0 to form the corresponding peptide hydrazide.
14 This allows ligation methods as described herein to
15 be performed at pH 6.0, without the need for a
16 potentially harmful thiol cofactor (useful if either
17 fragment or final construct is thiol sensitive) and
18 does not lead to the introduction of potentially
19 reactive side-chain groups (such as a thiol) into
20 the protein. Similarly, the inventors have
21 demonstrated that synthetic peptide C-terminal
22 thioesters specifically react with hydroxylamine
23 under aqueous conditions at pH 6.0 and pH 6.8 to
24 form the corresponding peptide hydroxamic acid.
25 In addition, as described below, the inventors have
26 demonstrated that both synthetic peptide C-terminal
27 thioesters and recombinant protein C-terminal
28 thioesters specifically react with O-
29 methylhydroxylamine under aqueous conditions at pH
30 7.5, to form the corresponding C-terminal N-methoxy
31 amide derivatives. This allows ligation methods as

1 described herein to be performed at pH 7.5, without
2 the need for a potentially harmful thiol cofactor.

3
4 Peptides and proteins that contain thioester
5 groups (where the peptide is the acyl substituent of
6 the thioester) can be reacted with hydrazine,
7 hydrazide or aminooxy derivatives of a label or a
8 peptide to afford site-specific labelling and
9 chemoselective ligation respectively (see, for
10 example, figures 4 and 5).

11
12 In an analogous fashion, peptides that contain
13 hydrazine, hydrazide or aminooxy groups can be
14 reacted with thioester derivatives of a label or a
15 peptide to afford site-specific labelling and
16 chemoselective ligation respectively (see, for
17 example, figures 4 and 5).

18
19 Furthermore, having demonstrated that recombinant
20 protein hydrazides can be generated by cleavage of
21 protein-intein fusions with hydrazine, the inventors
22 have shown that such protein hydrazides may be
23 ligated by reaction of the hydrazide moiety with
24 reactive groups other than activated ester moieties,
25 for example an aldehyde functionality or a ketone
26 functionality. For example, as described below, the
27 inventors have shown that a pyruvoyl derivative of a
28 synthetic peptide can be chemoselectively ligated to
29 the C-terminus of recombinant protein hydrazides
30 using the described approach, and in an analogous
31 fashion, a pyruvoyl derivative of fluorescein was
32 used to site-specifically label the C-terminus of

1 recombinant protein hydrazides using the described
2 approach.

3

4 This aspect of the invention provides a further
5 novel method of ligating a recombinant peptide to a
6 second peptide or indeed a label.

7

8 Thus, a ninth aspect of the invention provides a
9 method of producing an oligopeptide product, the
10 method comprising the steps:

- 11 a) providing a first oligopeptide, the first
12 oligopeptide having an aldehyde or ketone moiety,
- 13 b) providing a precursor oligopeptide molecule,
14 the precursor oligopeptide molecule comprising a
15 second oligopeptide fused N-terminally to an intein
16 domain,
- 17 c) reacting said precursor oligopeptide molecule
18 with hydrazine to generate an oligopeptide molecule
19 comprising an intermediate oligopeptide, said
20 intermediate oligopeptide having a C-terminal
21 hydrazide moiety,
- 22 d) allowing the aldehyde or ketone moiety of the
23 first oligopeptide to react with the hydrazide
24 moiety of the intermediate oligopeptide molecule to
25 form an oligopeptide product, in which first
26 oligopeptide and the second oligopeptide are linked
27 via a hydrazone linking moiety.

28

29 An example of this aspect is shown in Figure 6.

30

1 A tenth aspect of the invention provides a method of
2 labelling an oligopeptide, the method comprising the
3 steps:

4 a) providing a label molecule, the label molecule
5 having a aldehyde or ketone moiety,

6 b) providing a precursor oligopeptide molecule,
7 the precursor oligopeptide molecule comprising a
8 first oligopeptide fused N-terminally to an intein
9 domain,

10 c) reacting said precursor oligopeptide molecule
11 with hydrazine to generate an oligopeptide molecule
12 comprising an intermediate oligopeptide, said
13 intermediate oligopeptide having a terminal
14 hydrazide moiety,

15 d) allowing the aldehyde or ketone moiety of the
16 label molecule to react with the hydrazide moiety of
17 the intermediate oligopeptide molecule to form a
18 labelled oligopeptide product, in which the label
19 molecule and oligopeptide are linked via a hydrazone
20 linking moiety.

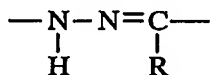
21

22 In preferred embodiments of the ninth and tenth
23 aspects of the invention, the hydrazone moiety has
24 Formula VII:

25

26

27



28

29 where R is H or any substituted or unsubstituted,
30 preferably unsubstituted, alkyl group.

31

1 In preferred aspects of the ninth and tenth aspects
2 of the invention, the method is performed at a pH in
3 the range pH 1.0 to pH 7.0, preferably pH 1.0 to pH
4 6.0, more preferably in the range pH 2.0 to pH 5.5,
5 most preferably in the range pH 2.0 to pH 4.5.

6
7 In a particular embodiment of the ninth and tenth
8 aspects of the invention, the aldehyde or ketone
9 containing moiety of the oligopeptide or of the
10 label is an α -diketone group or an α -keto aldehyde
11 group.

12
13 In a eleventh aspect of the present invention, there
14 is provided an oligopeptide product produced using a
15 method of the invention.

16
17 In an twelfth aspect, there is provided a labelled
18 oligopeptide comprising an oligopeptide labelled
19 according to a method of the invention.

20
21 Preferred features of each aspect of the invention
22 are as for each of the other aspects mutatis
23 mutandis.

24
25 The invention will now be described further in the
26 following non-limiting examples with reference made
27 to the accompanying drawings in which:

28
29 Figure 1 illustrates schematically the general
30 principle of chemical ligation.

31

1 Figure 2 illustrates schematically the mechanism of
2 protein splicing.

3

4 Figure 3 illustrates the generation of recombinant
5 C-terminal thioester proteins.

6

7 Figure 4 illustrates ligation of protein and peptide
8 thioesters with hydrazine and aminooxy containing
9 entities, such as labels, peptides and proteins.

10

11 Figure 5 illustrates the generation of synthetic and
12 recombinant peptide hydrazides for ligation with
13 thioester containing molecules. Note the peptide or
14 label is is the acyl substituent of the thioester.

15

16 Figure 6 illustrates the generation of recombinant
17 peptide hydrazides for ligation with aldehyde and
18 ketone containing molecules.

19

20 Figure 7 illustrates SDS-PAGE analysis of Grb2-SH2 -
21 GyrA - CBD (immobilised on chitin beads) treated
22 with DTT and MESNA. Molecular weight markers (lane
23 1); purified Grb2-SH2 - GyrA - CBD immobilised on
24 chitin beads (lane 4). Grb2-SH2 - GyrA - CBD treated
25 with 100 mM DTT (lanes 5 and 7) or 120 mM MESNA
26 (lanes 8 and 10). Both the whole reaction slurries
27 (lanes 5 and 8) and the reaction supernatants (lanes
28 7 and 10) were analysed.

29

30 Figure 8 illustrates SDS-PAGE analysis of Grb2-SH2 -
31 GyrA - CBD (immobilised on chitin beads) treated
32 with hydrazine. Molecular weight markers (lane 1);

1 Purified Grb2-SH2 - GyrA - CBD immobilised on chitin
2 beads after 20h treatment with phosphate buffer only
3 (lane 2). Grb2-SH2 - GyrA - CBD treated with 200 mM
4 hydrazine in phosphate buffer for 20 h. The whole
5 reaction slurries were analysed.

6

7 Figure 9 illustrates an ESMS spectrum of the C-
8 terminal hydrazide derivative of Grb2-SH2.

9

10 Figure 10 shows SDS-PAGE analysis of the reaction
11 between synthetic ketone containing peptide CH₃COCO-
12 myc with Grb2-SH2 - C-terminal hydrazide and
13 Cytochrome C. Molecular weight markers (lane 1);
14 Grb2-SH2 - C-terminal DTT thioester (lane 2).
15 Reaction between Grb2-SH2 - C-terminal hydrazide and
16 CH₃COCO-myc at time points t=0 h (lane 3), t=24 h
17 (lane 4), t= 48h (lane 5) and t= 72 h (lanes 6).
18 Reaction between Cytochrome C and CH₃COCO-myc at
19 time points t=0 h (lane 7), t=24 h (lane 8), t= 48h
20 (lane 9) and t= 72 h (lanes 10)

21

22 Figure 11 shows the structure of CH₃COCO-Lys(F1).
23 The 5-carboxy fluorescein positional isomer is
24 shown.

25

26 Figure 12 illustrates SDS-PAGE analysis of the
27 reaction between CH₃COCO-Lys(F1) with Grb2-SH2 C-
28 terminal hydrazide in 50 mM sodium acetate buffer pH
29 4.5. Molecular weight markers (lane 1); Grb2-SH2 C-
30 terminal hydrazide (lane 2). Reaction between Grb2-
31 SH2 C-terminal hydrazide and CH₃COCO-Lys(F1) at

1 time points t=4 h (lane 3), t=24 h (lane 4), t= 48h
2 (lane 5)

3

4 Figure 13 illustrates SDS-PAGE analysis of the
5 reaction between CH₃COCO-Lys(F1) with Cytochrome C
6 in 100 mM sodium acetate buffer pH 4.5. Molecular
7 weight markers (lane 1); Cytochrome C (lane 2).

8 Reaction between Cytochrome C and CH₃COCO-Lys(F1)
9 at time points t=4 h (lane 3), t=24 h (lane 4), t=
10 48h (lane 5).

11

12 Figure 14 illustrates SDS-PAGE analysis of the
13 reaction of CH₃COCO-Lys(F1) with Grb2-SH2 C-
14 terminal hydrazide and with Cytochrome C in 50 mM
15 sodium acetate buffer pH 4.5. (A) total protein stain
16 of gel. Prior to this coomassie staining (A), the
17 gel was imaged for green fluorescence (B). Molecular
18 weight markers (lane 1); Grb2 SH2 C-terminal
19 hydrazide (lane 2); Reaction between Grb2 SH2 C-
20 terminal hydrazide and CH₃COCO-Lys(F1) at time
21 points t=4 h (lane 3), t=24 h (lane 4), t= 48h (lane
22 5). Cytochrome C (lane 6); Reaction between
23 Cytochrome C and CH₃COCO-Lys(F1) at time points t=4
24 h (lane 7), t= 24 h (lane 8) and t= 48 h (lanes 9).

25

26 Figure 15 shows SDS-PAGE analysis of the reaction
27 between CH₃COCO-Lys(F1) and Grb2 SH2 C-terminal
28 hydrazide in 40% aqueous acetonitrile containing
29 0.1% TFA; reaction after 4 h (lane 1), 24 h (lane
30 2), 48h (lane 3), Grb2 SH2 C-terminal hydrazide
31 (lane 4).

32

1

2 **Examples**

3

4 **Example 1 -Protein ligation / site specific protein**
5 **labelling using the reaction of peptide / protein**
6 **thioesters with compounds containing hydrazine /**
7 **hydrazide or aminoxy functionalities.**

8

9 *A) Reaction of a peptide C-terminal thioester with*
10 *100mM hydrazine at pH 6.0*
11 200 mM sodium phosphate buffer pH 6.0 containing
12 100mM hydrazine monohydrate (200 µL) was added to a
13 model synthetic C-terminal thioester peptide termed
14 AS626p1A (200 µg) to yield a final peptide
15 concentration of 317 µM. AS626p1A has sequence ARTKQ
16 TARK(Me)₃ STGGKAPRKQ LATKAARK-COS-(CH₂)₂-COOC₂H₅ (SEQ
17 ID NO: 1) wherein a single Alanine residue (which
18 may be any one of the Alanine residues of SEQ ID NO:
19 1) is substituted by an Arginine residue. The
20 reaction was incubated at room temperature and
21 monitored with time by analytical reversed phase
22 HPLC. Vydac C18 column (5 µM, 0.46 x 25 cm). Linear
23 gradients of acetonitrile water / 0.1% TFA were used
24 to elute the peptides at a flow rate of 1 mL min⁻¹.
25 Individual peptides eluting from the column were
26 characterised by electrospray mass spectrometry.

27

28 *B)Reaction of a peptide C-terminal thioester with*
29 *100mM hydroxylamine at pH 6.0*
30 200 mM sodium phosphate buffer pH 6.0 containing
31 100mM hydroxylamine hydrogen chloride (200 µL) was

1 added to AS626p1A (200 µg) to yield a final peptide
2 concentration of 317 µM. The reaction was incubated
3 at room temperature and monitored with time by
4 analytical reversed phase HPLC. Vydac C18 column (5
5 µM, 0.46 x 25 cm). Linear gradients of acetonitrile
6 water / 0.1% TFA were used to elute the peptides at
7 a flow rate of 1 mL min⁻¹. Individual peptides
8 eluting from the column were characterised by
9 electrospray mass spectrometry.

10

11 *C) Reaction of a peptide C-terminal thioester with*
12 *100 mM hydroxylamine at pH 6.8*
13 200 mM sodium phosphate buffer pH 6.8 containing
14 100mM hydroxylamine hydrogen chloride (200 µL) was
15 added to AS626p1A (200 µg) to yield a final peptide
16 concentration of 317 µM. The reaction was incubated
17 at room temperature and monitored with time by
18 analytical reversed phase HPLC. Vydac C18 column (5
19 µM, 0.46 x 25 cm). Linear gradients of acetonitrile
20 water / 0.1% TFA were used to elute the peptides at
21 a flow rate of 1 mL min⁻¹. Individual peptides
22 eluting from the column were characterised by
23 electrospray mass spectrometry.

24

25 *D) Reaction of a peptide C-terminal thioester with*
26 *10mM hydroxylamine at pH 6.8*
27 The procedure as described in C) was repeated,
28 replacing 100mM hydroxylamine with 10mM
29 hydroxylamine.

30

1 *E) Reaction of a peptide C-terminal thioester with*
2 *10mM hydroxylamine at pH 7.5*

3 The procedure as described in D) was repeated, at
4 pH7.5.

5

6 *F) Reaction of a peptide C-terminal thioester with*
7 *2mM hydroxylamine at pH 7.5*

8 The procedure as described in E) was repeated,
9 replacing 10mM hydroxylamine with 2mM hydroxylamine.

10

11 *G) Reaction of a peptide C-terminal thioester with*
12 *100 mM O-Methylhydroxylamine (NH₂-O-CH₃) at pH 7.5*
13 200 mM sodium phosphate buffer pH 7.5 containing
14 100mM O-methylhydroxylamine (200 μ L) was added to
15 synthetic C-terminal thioester peptide AS626p1A (200
16 μ g) to yield a final peptide concentration of 317
17 μ M. The reaction was incubated at room temperature
18 and monitored with time by analytical reversed phase
19 HPLC. Vydac C18 column (5 μ M, 0.46 x 25 cm). Linear
20 gradients of acetonitrile water / 0.1% TFA were used
21 to elute the peptides at a flow rate of 1 mL min⁻¹.
22 Individual peptides eluting from the column were
23 characterised by electrospray mass spectrometry.

24

25 *H) Reaction of a peptide C-terminal thioester with*
26 *10 mM O-Methylhydroxylamine at pH 7.5*

27 The procedure as described in G) was repeated,
28 replacing 100 mM O-methylhydroxylamine with 10 mM O-
29 methylhydroxylamine.

30

1 *I) Reaction of a recombinant protein C-terminal*
2 *thioester with 100 mM O-Methylhydroxylamine at pH*
3 *7.5*

4
5 The C-terminal mercaptoethanesulfonic acid
6 thioester derivative of recombinant Grb2-SH2, was
7 generated through cleavage of the fusion protein
8 Grb2-SH2 - GyrA intein - CBD as described in Example
9 2 below. This recombinant C-terminal thioester
10 protein (100 µg) was reacted with 100mM O-
11 methylhydroxylamine in 200 mM sodium phosphate
12 buffer pH 7.5 (200 µL). The reaction was incubated
13 at room temperature and monitored with time by
14 analytical reversed phase HPLC. Vydac C5 column (5
15 µM, 0.46 x 25 cm). Linear gradients of acetonitrile
16 water / 0.1% TFA were used to elute the peptides at
17 a flow rate of 1 mL min⁻¹. Individual peptides
18 eluting from the column were characterised by
19 electrospray mass spectrometry.

20

21

22 **Results**

23 These examples demonstrate the novel strategy for
24 protein ligation / site specific protein labelling
25 of both synthetic and recombinant protein sequences
26 of the invention using the reaction of peptide /
27 protein C-terminal thioesters with compounds
28 containing hydrazine / hydrazide or aminoxy
29 functionalities.

30

1 As described above, a purified synthetic 27 amino
2 acid C-terminal thioester peptide (the ethyl 3-
3 mercaptopropionate thioester derivative) was treated
4 with hydrazine and hydroxylamine under various
5 conditions (Table 1).

6
7 Treatment with 100 mM hydrazine at pH 6.0 formed a
8 peptide species that eluted earlier than the
9 starting thioester peptide as analysed by HPLC. This
10 material was identified as the expected peptide
11 hydrazide by ESMS: observed mass = 3054 Da, expected
12 (av. isotope comp) 3053 Da. The reaction of the
13 peptide C-terminal thioester with hydrazine to form
14 the peptide hydrazide was monitored with time by
15 reverse phase HPLC. Only the desired material was
16 formed with no side product formation even after 3
17 days. The stability of the peptide hydrazide, under
18 the reaction conditions, indicates that the reaction
19 occurs at the C-terminal thioester moiety and is
20 chemoselective in nature. It also highlights the
21 applicability of this reaction for protein ligation
22 and labelling (2 h 70% conversion , 4h >95%
23 conversion).

24

25 To ascertain whether aminooxy containing compounds
26 chemoselectively react with peptide / protein C-
27 terminal thioesters, to afford protein ligation and
28 site-specific labelling, a synthetic C-terminal
29 thioester peptide was treated with hydroxylamine
30 under various conditions (Table 1).

31

1 A purified synthetic 27 amino acid C-terminal
2 thioester peptide (ethyl 3-mercaptopropionate
3 thioester, observed mass 3155 Da) was incubated at
4 room temperature with different hydroxylamine
5 concentrations in aqueous buffers of varying pH. In
6 all cases the peptide C-terminal thioester reacted
7 to form a single product that eluted earlier than
8 the starting thioester peptide as analysed by
9 reverse phase HPLC. This material corresponds to the
10 expected hydroxamic acid peptide as determined by
11 ESMS: observed mass = 3052 Da, expected (av. isotope
12 comp) 3054 Da. The kinetics of the reaction were
13 monitored using reverse phase HPLC. The peptide C-
14 terminal thioester was converted to the
15 corresponding peptide hydroxamic acid in a clean
16 fashion with no side-product formation. Increasing
17 the pH of the reaction buffer accelerated the rate
18 of reaction. For instance, with a concentration of
19 100mM NH_2OH , on moving from pH 6.0 to pH 6.8 the
20 percentage product formation after 1h increased from
21 25% to 91%. The rate of reaction with 100 mM NH_2OH
22 at pH 6.0, was comparable with 10 mM NH_2OH at pH 6.8.

23

24 The rate of reaction of the peptide C-terminal
25 thioester with hydroxalymine, to form the
26 corresponding hydroxamic acid, increases with
27 increasing pH and decreases with decreasing NH_2OH
28 concentrations. To identify conditions of pH and
29 reactant concentration suitable for peptide /
30 protein labelling and ligation, the labelling was
31 performed under increasing pH and decreasing NH_2OH
32 concentrations.

1
2 The reaction with 10 mM NH_2OH was 83% complete after
3 4h at pH 6.8, while at pH 7.5 it was 83% complete
4 after 2h. On further decreasing the NH_2OH
5 concentration to 2 mM the reaction rate at pH 7.5
6 decreased markedly, 70% of the starting peptide α -
7 thioester being converted to the corresponding
8 hydroxamic acid after 8hrs. It was noted that a
9 small amount of a side-product, corresponding in
10 mass to the peptide acid, was formed during the
11 reaction. Presumably this was formed by a competing
12 hydrolysis side reaction at pH 7.5, which was not
13 observed with 10 mM NH_2OH at pH 7.5 due to the
14 faster reaction at this higher reactant
15 concentration.

16

17

Reactant	Concent ration	pH	Percentage product formation with time				
			1hr	2hr	4hr	8hr	72hr
NH_2NH_2	100 mM	6.0	-	70	100		
NH_2OH	100 mM	6.0	25	48.1	76.3	-	100
NH_2OH	100 mM	6.8	91	100			
NH_2OH	10 mM	6.8	26	-	83	100	
NH_2OH	10 mM	7.5	-	82.7	100	100	
NH_2OH	2 mM	7.5	11.2	17	38	70	80*

18 **Table 1**

19 *All starting material has reacted with 80%
20 conversion to the desired product and ~20% to the
21 hydrolysis side-product.

22

23 To further investigate the chemoselective reaction
24 of aminoxy containing compounds with peptide /

1 protein C-terminal thioesters, to afford protein
2 ligation and site-specific labelling, the synthetic
3 C-terminal thioester peptide AS626p1 was treated
4 with *O*-methylhydroxylamine.

5 The purified synthetic 27 amino acid C-terminal
6 thioester peptide (ethyl 3-mercaptopropionate
7 thioester, observed mass 3155 Da) was incubated at
8 room temperature with 100mM *O*-methylhydroxylamine in
9 200 mM sodium phosphate buffer pH 7.5. The peptide
10 C-terminal thioester reacted to form a single
11 product that eluted earlier than the starting
12 thioester peptide as analysed by reverse phase HPLC.
13 This material corresponded to the expected N-methoxy
14 peptide amide as determined by ESMS: observed mass =
15 3070 Da, expected mass 3068 Da. The kinetics of the
16 reaction were monitored using reverse phase HPLC
17 (Table II). The peptide C-terminal thioester was
18 converted to the corresponding N-methoxy peptide
19 amide derivative in a clean fashion with no side-
20 product formation, with the reaction 75% complete
21 after 24 h. Under these conditions no thioester
22 hydrolysis was observed.

23

Reactant	Concentration	pH	Percentage product formation with time				
			1hr	2hr	5hr	24hr	72hr

NH ₂ OCH ₃	100 mM	7.5	-	7.5	28	76	
----------------------------------	--------	-----	---	-----	----	----	--

1 Table II

2

3 When the reaction was repeated under the same
4 conditions but with 10 mM *O*-methylhydroxylamine
5 replacing 100 mM *O*-methylhydroxylamine, the reaction
6 rate was slower. However, after 72h, 88% of the
7 starting C-terminal thioester peptide had reacted.
8 Under these conditions side-product formation was
9 observed, in addition to the desired reaction
10 product formation. Even so, after 72h, 30-40% of the
11 reaction product was estimated to be the desired
12 ligation reaction product (N-methoxy peptide amide)
13 from HPLC analysis of the reaction mixture.

14

15 The reaction of *O*-methylhydroxylamine with
16 recombinant C-terminal thioester proteins was also
17 investigated. Recombinant Grb2-SH2 was generated as
18 the C-terminal mercaptoethanesulfonic acid
19 thioester derivative, through thiol mediated
20 cleavage of the fusion protein Grb2-SH2 - GyrA
21 intein - CBD, as described in Example 2. This
22 recombinant C-terminal thioester protein was reacted
23 with 100mM *O*-methylhydroxylamine at pH 7.5. Analysis
24 of the reaction mixture after 18h by HPLC and ESMS
25 showed that all of the C-terminal thioester protein

1 had been completely converted into two protein
2 species. These two protein derivatives corresponded
3 to the desired ligation reaction product, namely
4 Grb2-SH2 C-terminal N-methoxy amide (expected mass
5 12067 Da; observed mass 12067 Da), and an oxidised
6 form of the desired reaction product (observed mass
7 12084 Da). No side products corresponding to
8 hydrolysis of the C-terminal protein thioester were
9 observed. Thus all of the C-terminal thioester
10 recombinant protein had chemoselectively ligated
11 with *O*-hydroxylamine, via an amide bond forming
12 reaction specifically at the C-terminus of the
13 protein. i.e. the reaction afforded site-specific C-
14 terminal labelling of the recombinant protein.

15

16

17

18 **Example 2- Generation of recombinant C-terminal**
19 **hydrazide Grb2 SH2 protein.**

20

21 To investigate (i) the ability to generate
22 recombinant C-terminal hydrazide proteins through
23 the selective cleavage of protein - intein fusions
24 with hydrazine, and (ii) their subsequent use in
25 ligation / labelling reactions, the SH2 domain of
26 the adapter protein Grb2 was chosen as a model
27 system.

28

29 Sequence of human Grb2 SH2 domain

1 HPW FFGKIPRAKA EEMLSKQRHD GAFLIRESES APGDFSLSVK
2 FGNDVQHFVKV LRDGAGKYFL WVKFNSLNE LVDYHRSTSV
3 SRNQQIFLRD IEQVPQQPT
4

5 *Expression of Grb2-SH2 domain - GyrA intein fusion.*
6 The DNA sequence encoding the SH2 domain of human
7 Grb2 appended at its C-terminus with an extra
8 glycine residue was cloned into the pTXB1 expression
9 plasmid (NEB). This vector pTXB1_{Grb2-SH2 (Gly)} encodes
10 for a fusion protein whereby the SH2 domain of Grb2
11 is linked via a glycine residue to the N-terminus of
12 the GyrA intein, which is in turn fused to the N-
13 terminus of a chitin binding domain region (CBD).
14 *E. coli* cells were transformed with this plasmid and
15 grown in LB medium to mid log phase and protein
16 expression induced for 4h at 37°C with 0.5 mM IPTG.
17 After centrifugation the cells were re-suspended in
18 lysis buffer (0.1 mM EDTA, 250 mM NaCl, 5% glycerol,
19 1 mM PMSF, 25 mM HEPES, pH 7.4) and lysed by
20 sonication. The soluble fraction was loaded onto a
21 chitin column pre-equilibrated in lysis buffer. The
22 column was then washed with wash buffer (1 mM EDTA,
23 250 mM NaCl, 0.1% Triton-X 100, , 25 mM HEPES, pH
24 7.0) to yield purified Grb2-SH2 - GyrA-CBD
25 immobilised on chitin beads (Figure 7).
26

27 *Generation of Grb2-SH2 C-terminal thioesters by*
28 *thiol induced cleavage of the Grb2-SH2 - GyrA intein*
29 *fusion.*

30 To ascertain that the intein domain within the
31 protein was functional the fusion protein was
32 exposed to thiols to assess the extent of cleavage

1 via transthioesterification. Chitin beads containing
2 immobilised Grb2-SH2 - GyrA-CBD were equilibrated
3 into 200 mM NaCl, 200 mM phosphate buffer pH 7.4.
4 Dithiothreitol (DTT) or 2-mercaptoethanesulfonic
5 acid (MESNA) were then added to the beads in 200 mM
6 NaCl, 200 mM phosphate buffer pH 7.4 to give a 50%
7 slurry with a final thiol concentration of 100 mM or
8 120 mM respectively. The mixtures were then rocked
9 at room temperature and aliquots analysed by SDS-
10 PAGE. After 48 hours the supernatants from the
11 reactions were isolated and subsequently analysed by
12 HPLC and ESMS.

13
14 Treatment of Grb2-SH2 - GyrA intein - CBD fusion
15 with both DTT and MESNA resulted in cleavage of the
16 fusion protein into two protein species (Figure 7).
17 The molecular size of the two fragments corresponds
18 to that of the Grb2 - SH2 and the GyrA - intein
19 fusion, indicative that cleavage has taken place at
20 the SH2 - intein junction. Cleavage of the precursor
21 fusion protein liberated the SH2 domain into the
22 supernatant while the GyrA intein-CBD portion
23 remained immobilized on the chitin beads. After
24 cleavage with both DTT or MESNA, ESMS analysis of
25 the supernatants confirmed that the Grb2-SH2 was
26 generated as either the expected DTT or MESNA C-
27 terminal thioester derivatives respectively.

28
29 Expected mass of Grb2-SH2 DTT - C-terminal thioester
30 = 12173.9 Da; observed mass 12173.5 Da. Expected
31 mass of Grb2-SH2 MESNA - C-terminal thioester =
32 12162.0 Da; observed mass 12163.0 Da.

1
2 *Generation of Grb2-SH2 C-terminal hydrazide by*
3 *hydrazine induced cleavage of the Grb2-SH2 - GyrA*
4 *intein fusion.*

5
6 The inventors hypothesised that the thioester
7 linkage between Grb2-SH2 and the GyrA intein in the
8 precursor fusion protein is cleaved with hydrazine.
9 The chemoselective reaction of hydrazine, at the
10 thioester moiety linking Grb2 SH2 to the intein,
11 would liberate the Grb2-SH2 domain into the
12 supernatant as its corresponding C-terminal
13 hydrazide derivative. Chitin beads containing
14 immobilised Grb2-SH2 - GyrA-CBD were therefore
15 equilibrated into 200 mM NaCl, 200 mM phosphate
16 buffer pH 7.4 and hydrazine monohydrate added in the
17 same buffer to give a 50% slurry with a final
18 hydrazine concentration of 200 mM. The mixture was
19 then rocked at room temperature and analysed by SDS-
20 PAGE (Figure 8). After 20 hours the supernatant was
21 removed and analysed by HPLC and ESMS.

22
23 Treatment of Grb2-SH2 - GyrA intein - CBD fusion
24 with hydrazine resulted in cleavage of the fusion
25 protein into two species. The molecular size of the
26 two fragments as analysed by SDS-PAGE corresponded
27 to Grb2 - SH2 and the GyrA - intein fusion,
28 indicative that cleavage has taken place at the
29 unique thioester linkage between the SH2 and intein
30 domains. Cleavage of the precursor fusion protein
31 liberated the SH2 domain into the supernatant while
32 the GyrA intein-CBD portion remained immobilized on

1 the chitin beads. HPLC and ESMS analysis of the
2 cleavage supernatant confirmed that a single protein
3 species was generated that corresponds to the C-
4 terminal hydrazide derivative of Grb2-SH2. Expected
5 mass of Grb2-SH2 C-terminal hydrazide = 12051.7 Da;
6 observed mass 12053.0 Da. (Figure 9)

7
8 After 20 h of reaction Grb2-SH2 C-terminal hydrazide
9 was isolated from the supernatant by either (i)
10 using RPHPLC followed by lyophilisation or (ii) by
11 gel filtration. In this later approach the Grb2-SH2
12 C-terminal hydrazide reaction solution was loaded
13 onto a superdex peptide column (Amersham
14 Biosciences) and eluted with a running buffer of 50
15 mM sodium acetate pH 4.5. This yielded a solution of
16 purified Grb2-SH2 C-terminal hydrazide in 50 mM
17 sodium acetate pH 4.5. This solution was
18 concentrated using a centricon filter (3000 MWCO),
19 then snap frozen and stored at -20°C until use.

20
21 A sample of the purified and lyophilised Grb2-SH2 C-
22 terminal hydrazide (100 µg) was treated with the
23 protease Lys-C (5 µg) in 100mM ammonium bicarbonate
24 buffer pH 8.2 (100 µL). After incubating at 30°C
25 overnight the reaction was lyophilised and analysed
26 by MALDI mass spectrometry. The observed mass of the
27 C-terminal proteolytic fragment
28 (FNSLNELVDYHRSTSVSRNQQIFLRDIEQVPQQPTG) corresponds
29 to that of the desired C-terminal hydrazide

1 derivative (expected mass of C-terminal hydrazide
2 proteolytic fragment 4229 Da; observed mass 4231
3 Da)
4
5

6 **Example 3- Generation of recombinant C-terminal**
7 **hydrazide maltose binding protein.**
8

9 As a further demonstration of the described
10 approach, for generating recombinant C-terminal
11 hydrazide proteins through the selective cleavage of
12 protein - intein fusions with hydrazine, the
13 generation of the C-terminal hydrazide derivative of
14 maltose binding protein (MBP) was investigated.
15

16 Sequence of human MBP used

17 M K I E E G K L V I W I N G D K G Y N G L A E V G K
18 K F E K D T G I K V T V E H P D K L E E K F P Q V A
19 A T G D G P D I I F W A H D R F G G Y A Q S G L L A
20 E I T P D K A F Q D K L Y P F T W D A V R Y N G K L
21 I A Y P I A V E A L S L I Y N K D L L P N P P K T W
22 E E I P A L D K E L K A K G K S A L M F N L Q E P Y
23 F T W P L I A A D G G Y A F K Y E N G K Y D I K D V
24 G V D N A G A K A G L T F L V D L I K N K H M N A D
25 T D Y S I A E A A F N K G E T A M T I N G P W A W S
26 N I D T S K V N Y G V T V L P T F K G Q P S K P F V
27 G V L S A G I N A A S P N K E L A K E F L E N Y L L
28 T D E G L E A V N K D K P L G A V A L K S Y E E E L
29 A K D P R I A A T M E N A Q K G E I M P N I P Q M S
30 A F W Y A V R T A V I N A A S G R Q T V D E A L K D

1 A Q T N S S S N N N N N N N N N N L G I E G R G T L
2 E G

3

4

5 *Expression of MBP - Sce VMA intein fusion.*

6 The expression vector pMYB5 (New England Biolabs)
7 encodes for a fusion protein comprising maltose
8 binding protein (sequence above) fused N-terminal to
9 the *Sce* VMA intein, which is in turn fused to the N-
10 terminus of a chitin binding domain (CBD) to
11 facilitate purification.

12

13 *E. coli* cells were transformed with this plasmid and
14 grown in LB medium to mid log phase and protein
15 expression induced for 4h at 37°C with 0.5 mM IPTG.
16 After centrifugation the cells were re-suspended in
17 lysis buffer (0.1 mM EDTA, 250 mM NaCl, 5% glycerol,
18 1 mM PMSF, 25 mM HEPES, pH 7.4) and lysed by
19 sonication. The soluble fraction was loaded onto a
20 chitin column pre- equilibrated in lysis buffer. The
21 column was then washed with wash buffer (1 mM EDTA,
22 250 mM NaCl, 0.1% Triton-X 100, , 25 mM HEPES, pH
23 7.0) to yield the purified fusion protein (MBP-VMA-
24 CBD) immobilised on chitin beads.

25

26 *Generation of MBP C-terminal thioesters by thiol*
27 *induced cleavage of the MBP - VMA- intein fusion*
28 *protein.*

29

30 To ascertain that the intein domain within MBP-VMA-
31 CBD was functional, the fusion protein was exposed
32 to 2-mercaptoethanesulfonic acid (MESNA) to assess

1 the extent of cleavage via transthioesterification.
2 Chitin beads containing immobilised MBP-VMA-CBD were
3 equilibrated into 200 mM NaCl, 200 mM phosphate
4 buffer pH 7.4. MESNA was then added to the beads in
5 200 mM NaCl, 200 mM phosphate buffer pH 7.4 to give
6 a 50% slurry with a final thiol concentration of 120
7 mM. The mixture was then rocked at room temperature
8 and aliquots analysed by SDS-PAGE. After 48 hours
9 the supernatants from the reactions were isolated
10 and subsequently analysed by HPLC and ESMS.

11

12 Treatment of MBP-VMA-CBD fusion with MESNA results
13 in cleavage of the fusion protein into two protein
14 species. The molecular size of the two fragments
15 corresponds to that of the MBP and the VMA-CBD
16 portion, indicative that cleavage has taken place at
17 the MBP - VMA intein junction. Cleavage of the
18 precursor fusion protein liberates MBP into the
19 supernatant while the VMA-CBD portion remains
20 immobilized on the chitin beads. This was confirmed
21 by ESMS analysis of the cleavage supernatant, which
22 contained one protein species. Expected mass of MBP
23 C-terminal MESNA thioester 43064 Da; observed mass
24 43098 Da.

25

26 *Generation of MBP C-terminal hydrazide by hydrazine*
27 *induced cleavage of the MBP-VMA intein fusion*
28 *protein.*

29

30 Chitin beads containing immobilised MBP-VMA-CBD were
31 equilibrated into 200 mM NaCl, 200 mM phosphate
32 buffer pH 7.4 and hydrazine monohydrate added in the

1 same buffer to give a 50% slurry with a final
2 hydrazine concentration of 200 mM. The mixture was
3 then rocked at room temperature and analysed by SDS-
4 PAGE and by HPLC and ESMS.

5
6 After 20 h of reaction MBP C-terminal hydrazide was
7 isolated from the supernatant by either (i) using
8 RPHPLC followed by lyophilisation or (ii) by gel
9 filtration. In this later approach the MBP C-
10 terminal hydrazide reaction solution was loaded onto
11 a superdex peptide column (Amersham Biosciences) and
12 eluted with a running buffer of 50 mM sodium acetate
13 buffer pH 4.5. This yielded a solution of purified
14 MBP C-terminal hydrazide in 50 mM sodium acetate
15 buffer pH 4.5. This protein solution was
16 concentrated using a centricon filter (3000 MWCO),
17 then snap frozen and stored at -20°C until use.

18
19 Treatment of MBP-VMA-CBD fusion with hydrazine
20 results in cleavage of the fusion protein into two
21 species. The molecular size of the two fragments as
22 analysed by SDS-PAGE corresponds to MBP and the VMA-
23 CBD portion, indicative that cleavage has taken
24 place at the unique thioester linkage between the
25 MBP - VMA intein domain. Cleavage of the precursor
26 fusion protein liberates MBP into the supernatant,
27 while the VMA-CBD portion remains immobilized on the
28 chitin beads. HPLC and ESMS analysis of the cleavage
29 supernatant confirms that a single protein species
30 is generated with an observed mass of 42988 Da. The
31 expected mass difference between the C-terminal
32 MESNA thioester derivative of a protein and its

1 corresponding C-terminal hydrazide is 111 Da. The
2 observed mass of the C-terminal MESNA thioester of
3 MBP was found to be 43098 Da. Thus the product from
4 the hydrazine cleavage of MBP-VMA- CBD is 110 Da
5 lower, indicating that the desired C-terminal
6 hydrazide derivative of MBP had been formed.

7

8 **Example 4- Ligation of aldehyde and ketone**
9 **containing peptides and labels to recombinant C-**
10 **terminal hydrazide containing proteins: Ligation of**
11 **a synthetic peptide c-myc to recombinant Grb2 SH2**
12 **domain.**

13

14 The inventors hypothesised that recombinant protein
15 C-terminal hydrazides, generated by hydrazine
16 treatment of the corresponding intein fusion
17 precursor, can be site-specifically modified by
18 chemoselective ligation with aldehyde and ketone
19 containing peptides and labels. To demonstrate such
20 an approach, the ability of a synthetic ketone
21 containing peptide to ligate with the Grb2-SH2 C-
22 terminal hydrazide generated above was investigated.
23 A synthetic peptide corresponding to the c-myc
24 epitope sequence was synthesised GEQKLISEEDL-NH₂,
25 whereby pyruvic acid was coupled to the amino
26 terminus of the peptide as the last step of the
27 assembly. This peptide (designated CH₃COCO-myc) was
28 purified to > 95% purity by RPHPLC and lyophilised
29 (ESMS expected monoisotopic mass 1328.6 Da; observed
30 mass 1328.6 Da).

31

1 A sample of CH₃COCO-myc peptide was dissolved in 100
2 mM sodium acetate buffer pH 4.5 to give a 4 mM
3 peptide concentration. This peptide solution (100
4 µL) was then added to an aliquot of lyophilised
5 Grb2-SH2 C-terminal hydrazide protein (~ 250 µg) and
6 the reaction monitored by SDS-PAGE (Figure 10) As a
7 control CH₃COCO-myc was also incubated with
8 Cytochrome C, a protein of similar same size to
9 Grb2-SH2 but absent of a hydrazide functionality.
10
11 SDS-PAGE analysis shows that CH₃COCO-myc peptide
12 has indeed ligated with Grb2-SH2 C-terminal
13 hydrazide, as indicated by the conversion of Grb2-
14 SH2 C-terminal hydrazide into a protein species of
15 a higher molecular weight (approximately 1000-2000
16 Da higher). The reaction is virtually complete after
17 24 h and the reaction product appears to be stable.
18 On the other hand, there was no observable change to
19 Cytochrome C with time i.e no ligation, establishing
20 that the ligation reaction is occurring at the C-
21 terminal hydrazide functionality of Grb2-SH2.
22
23 After 96 h of reaction the product from the Grb2-SH2
24 ligation reaction was isolated by HPLC and
25 characterised by ESMS. Chemoselective ligation of
26 CH₃COCO-myc to Grb2-SH2 C-terminal hydrazide via
27 hydrazone bond formation would give a product of
28 expected mass 13363.7 Da. The observed product mass
29 was 13364.1 Da indicting that the desired ligation
30 product had been formed.
31

1 **Example 5- Ligation of aldehyde and ketone**
2 **containing peptides and labels to recombinant C-**
3 **terminal hydrazide containing proteins: Fluorescein**
4 **labelling of Grb2-SH2.**

5
6 In this example the recombinant C-terminal hydrazide
7 derivative of Grb2-SH2, generated through hydrazine
8 cleavage of the precursor intein fusion protein, was
9 reacted with a ketone containing derivative of
10 fluorescein to afford site-specific fluorescent
11 labelling of the protein.

12
13 To facilitate fluorescent labelling of C-terminal
14 hydrazide recombinant proteins using the described
15 approach, the fluorophore needs to contain the
16 appropriate reactive group for ligation, namely an
17 aldehyde or ketone functionality. To this end a
18 derivative of fluorescein was synthesized containing
19 a pyruvoyl moiety. Initially, Fmoc-Lys(Mtt)-OH was
20 coupled to a rink amide resin, and the Mtt group
21 removed using standard procedures (1% TFA, 4%
22 triisopropylsilane in dichloromethane). 5(6)-
23 carboxyfluorescein was then couple to the lysine ϵ -
24 amino group. The Fmoc group was then removed and
25 pyruvic acid coupled to the free α -amino group of
26 the lysine. After cleavage from the resin, the
27 desired fluorescein derivative [designated CH₃COCO-
28 Lys(Fl), see Figure 11] was purified to > 95% purity
29 by RPHPLC and lyophilised (ESMS, expected
30 monoisotopic mass 576.2 Da; observed monoisotopic
31 mass 576.0 Da).

32

1 To establish the reactivity of CH₃COCO-Lys(F1) with
2 C-terminal hydrazide peptides and proteins, the
3 reaction of CH₃COCO-Lys(F1) with a small synthetic
4 C-terminal hydrazide peptide SLAYG-NHNH₂ was
5 investigated. A sample of CH₃COCO-Lys(F1) and SLAYG-
6 NHNH₂ peptide were co-dissolved in 100 mM sodium
7 acetate buffer pH 4.5 to give final concentrations
8 of 0.3 mM and 2 mM respectively. After 20 h
9 incubation at room temperature, the reaction was
10 deemed complete as determined by RPHPLC analysis.
11 All the starting CH₃COCO-Lys(F1) had reacted to give
12 predominantly a single product. The mass of which
13 corresponds to the desired ligation product, namely
14 conjugation of the two reactants via hydrazone bond
15 formation (ESMS expected monoisotopic mass 1079 Da;
16 observed mass 1080 Da).

17

18 Having established the specific reaction of CH₃COCO-
19 Lys(F1) with hydrazide containing peptides, this
20 fluorescein derivative was used for the site-
21 specific labeling of recombinant Grb2 SH2 C-terminal
22 hydrazide (generated through hydrazine cleavage of
23 Grb2 SH2 - GyrA - CBD).

24

25 Two complementary methods were employed for the
26 purification of Grb2 SH2 C-terminal hydrazide from
27 the fusion protein cleavage reaction (Example 2).
28 The purified protein was isolated as either a
29 lyophilized solid or in a solution of 50 mM sodium
30 acetate buffer pH 4.5. This latter buffer system was
31 chosen as the pH is suited to hydrazone bond forming
32 reactions. An aliquot of Grb2 SH2 C-terminal

1 hydrazide in 50mM sodium acetate pH 4.5 (250 µg, 200
2 µL) was added directly to a sample of CH₃COCO-
3 Lys(Fl) to give a final concentration of fluorophore
4 of circa 0.3 mM. The reaction was incubated at room
5 temperature and monitored by SDS-PAGE. As a control
6 CH₃COCO-Lys(Fl) was also incubated under the same
7 conditions with Cytochrome C, a protein of similar
8 same size to Grb2-SH2 but absent of a hydrazide
9 functionality.

10

11 SDS-PAGE analysis shows that CH₃COCO-Lys(Fl) has
12 indeed ligated with Grb2-SH2 C-terminal hydrazide
13 (Figure 12) as indicated by the conversion of Grb2-
14 SH2 C-terminal hydrazide into a single protein
15 species with an apparent increase in molecular
16 weight (approximately 1000-2000 Da higher). After
17 SDS-PAGE analysis of the reactions, fluorescence
18 imaging of the gel confirmed that the newly formed
19 reaction product contains a fluorescein label, and
20 that the reaction is clean, with only a single
21 fluorescent protein product being formed (figure
22 14). The reaction is virtually complete after 24 h
23 and the reaction product appears to be stable under
24 these conditions.

25

26 On the other hand there was no observable change to
27 Cytochrome C over the time course of the experiment
28 i.e no ligation (Figure 13) with a complete absence
29 of the formation of any fluorescent protein products
30 (Figure 14). Thus establishing that the ligation
31 reaction is occurring at the C-terminal hydrazide
32 functionality of Grb2 SH2, to yield site-specific C-

1 terminal fluorescent labelling of the recombinant
2 protein. After 48 h of reaction, the product from
3 the ligation reaction with Grb2 SH2 was isolated by
4 HPLC. The mass of this product, by ESMS, confirmed
5 the addition of one fluorescein group to the
6 protein.

7
8 In another example, lyophilised Grb2 SH2 C-terminal
9 hydrazide was directly dissolved into 100 mM sodium
10 acetate pH 4.5 and added to CH₃COCO-Lys(F1). Whilst
11 some protein precipitation was observed, the soluble
12 fraction of the protein reacted with CH₃COCO-Lys(F1)
13 in the anticipated manner described above.

14
15 In an alternative strategy, a lyophilized sample of
16 Grb2 SH2 C-terminal hydrazide (250 µg) was dissolved
17 in 40% aqueous acetonitrile containing 0.1% TFA
18 (200 µL). This solution was then added to a sample
19 of CH₃COCO-Lys(F1) to give a final fluorophore
20 concentration of circa 0.3 mM. The solution was
21 incubated at room temperature and the reaction
22 periodically analyzed. SDS-PAGE analysis showed that
23 the labeling reaction had occurred cleanly and
24 rapidly under these conditions (Figure 15). Grb2 SH2
25 C-terminal hydrazide was converted into a single
26 protein species with an apparent increased molecular
27 weight expected for that of the desired product, and
28 this newly formed protein was green fluorescent when
29 visualised under a UV lamp. ESMS of the reaction
30 product confirmed that one fluorescein molecule had
31 been added to the protein. The reaction is virtually
32 complete after 4 h, with prolonged incubation

1 appearing to be detrimental to the formation of the
2 ligation product.

3

4 **Example 6- Ligation of aldehyde and ketone**
5 **containing peptides and labels to recombinant C-**
6 **terminal hydrazide containing proteins: Fluorescein**
7 **labelling of MBP.**

8

9 As a further exemplification, the described approach
10 was used for the site-specific C-terminal labeling
11 of MBP with fluorescein. A sample (250 µg) of
12 lyophilised recombinant MBP C-terminal hydrazide
13 (generated through hydrazine cleavage of MBP - VMA -
14 CBD precursor fusion protein) was dissolved in 40%
15 aqueous acetonitrile containing 0.1% TFA (200 µL).
16 The solution was then added to a sample of CH₃COCO-
17 Lys(Fl) to give a final fluorophore concentration of
18 circa 0.3 mM. The reaction was then incubated at
19 room temperature and periodically analyzed by SDS-
20 PAGE.

21

22 SDS-PAGE analysis showed that the fluorescein
23 labelling reaction had occurred under these
24 conditions, as indicated by the formation of a
25 single green fluorescent species with a molecular
26 weight of circa 42 KDa. MALDI analysis of the
27 reaction mixture after 48 h was consistent with the
28 addition of one fluorescein molecule to MBP.

29

30 In summary, the present invention provides novel
31 methods of protein ligation and protein labelling.
32 These enable both synthetic and recombinantly

1 derived protein fragments to be efficiently joined
2 together in a regioselective manner. This thus
3 enables large proteins to be constructed from
4 combinations of synthetic and recombinant fragments
5 and allows proteins of any size to be site-
6 specifically modified in an unprecedented manner.
7 This is of major importance for biological and
8 biomedical science and drug discovery when one
9 considers that the ~ 30,000 human genes yield
10 hundreds of thousands of different protein species
11 through post-translational modification. Such post-
12 translationally modified proteins cannot be accessed
13 through current recombinant technologies.

14
15 The application of such protein ligation techniques
16 may be used for protein based tools, protein
17 therapeutics and in de novo design and may open up
18 many new avenues in biological and biomedical
19 sciences that have hitherto not been possible.

20
21 All documents referred to in this specification are
22 herein incorporated by reference. Various
23 modifications and variations to the described
24 embodiments of the inventions will be apparent to
25 those skilled in the art without departing from the
26 scope and spirit of the invention. Although the
27 invention has been described in connection with
28 specific preferred embodiments, it should be
29 understood that the invention as claimed should not
30 be unduly limited to such specific embodiments.
31 Indeed, various modifications of the described modes
32 of carrying out the invention which are obvious to

1 those skilled in the art are intended to be covered
2 by the present invention.
3
4